# Magnetic resonance spectroscopy at 3 T for *in vivo* detection of 2-hydroxyglutarate in lower grade gliomas (LGG)

E. Anghileri<sup>1</sup>, N. Bertolino<sup>2</sup>, V. Cuccarini<sup>2</sup>, I. Zucca<sup>2</sup>, Gaetano Finocchiaro<sup>1</sup>, M. Eoli<sup>1</sup>, G. Tringali<sup>3</sup>, F. Di Meco<sup>3</sup>, P. Ferroli<sup>3</sup>, MG Bruzzone<sup>2</sup>

<sup>1</sup>Unit of Molecular Neuro-oncology, <sup>2</sup>Unit of Neuroradiology, <sup>3</sup>Neurosurgery Department, IRCCCS C.Besta, Via Celoria 11, 20133 Milan, Italy. <u>e-mail contact</u>: anghileri.e@istituto-besta.it

#### Introduction

Lower grade gliomas (LGG) are rare brain tumor, characterized by isocitrate dehydrogenase 1/2 (IDH1/2) mutations that induce 2-hydroxyglutarate (2HG) production [1]. Hydrogen magnetic-resonance spectroscopy (1H-MRS) can be used to detect 2HG [2-4], although such metabolite overlaps with N-acetylaspartate (NAA), glutamate (Glu), glutamine (Gln) and GABA spectra at 3 Tesla spectral resolution. We tested in a previous work that the concentration of 2HG would be partially but significantly confounded by the concentration of NAA and Glu molecules on phantoms (Fig.1) [5]. We move on investigating the feasibility of 2HG detection *in vivo* LGG patients by 1H-MRS, associated to the opportune sequence configuration. We will detect 2HG in *ex vivo* tumoral samples by liquid chromatography tandem mass spectrometry (LC-MS/MS) to support our data. Such approach will be useful to identify IDH1/2 mutant LGG by non-invasive means, helping early diagnosis and furnishing a dynamic tool, also considering the



#### Fig. 1

Effect of NAA and Glu concentration on 2HG measurements. The charts represent absolute 2HG concentration error with respect

**Methods** Spectra from LGG patients were respectively acquired with a 1H-MRS single-voxel PRESS sequence with TE=30 ms (group 1) and a tailored PRESS sequence with TE=97 ms (group 2). The MR protocol included two MRS acquisition for each subject: a voxel within the tumor and one contralateral. The voxel size was 2x1.5x1.5 cm<sup>3</sup> for group 1 and group 2A (n=14 total) and 2x2x2 cm<sup>3</sup> for group 2B (n=19 total). Group 1 counts 12 patients, group 2 (2A+2B) 33 patients. MR 3T scanner with a 32 channel head coil was used. Metabolite concentrations were estimated by linear combination analysis and a simulated basis set using LCModel software. IDH1/2 analysis were performed by immunochemistry and/or PCR.



**Fig. 2** An example of sagittal T1 (a), axial T2 (b) and coronal (c) FLAIR plains at TE 97 ms, with acquisition voxel = 2x2x2 cm<sup>3</sup>.

#### **Tab. 1** Specificy and sensitivity among the groups.

$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	TE 30	IDH1 mut	IDH1 wt	TE 97 small	IDH1 mut	IDH1 wt	TE 97 large	IDH1 mut	IDH1 wt
2HG- MRS-       4       2       2HG- MRS-       6       2       2HG- MRS-       4       6         tot       10       2       tot       10       4       tot       13       6	2HG- MRS+	6	0	2HG- MRS+	4	2	2HG- MRS+	9	0
tot 10 2 tot 10 4 tot 13 6	2HG- MRS-	4	2	2HG- MRS-	6	2	2HG- MRS-	4	6
	tot	10	2	tot	10	4	tot	13	6

#### Results

Spectra at TE=30 showed low specificity: 2 false positive (FP) and 4 false negative (FN) were identified out of the 12 LGG patients (10 IDH1mut and 2 IDH1wt). Group 2A correctly identified 4 mutated and 2 wt LGG, but also 2 FP and 6 FN out of the 14 patients (10 IDH1mut and 4 IDH1 wt). The larger voxel TE97 sequence resulted in higher specificity: no FP and 4 FN only (including two with technical issues: one suboptimal spectrum and one including 40% normal tissue) were detected (Tab.1). Besides, TE30 sequence detected 2HG in the contralateral region in 5 cases, while TE=97 sequences did not (Fig. 2, 3).





#### Conclusions

Our data suggest that MRS TE97 is rather specific for 2HG detection and the 2x2x2 cm<sup>3</sup> voxel bring to highest specificity and sensitivity than a smaller one. Larger number of patients will confirm the accuracy of the technique. We will also pair the *in vivo* detection by MRS with 2HG quantification by HPLC on tumoral samples and other biological fluids. The detection of 2HG could help for a non-invasive diagnosis, clinical follow-up, and possibly patient screening for targeted anti-IDH1mut therapy.

**Fig. 4** Examples of spectra analysis from the three PRESS sequences configuration: short TE (30 ms) spectrum (a), longer TE (97 ms) spectrum (b) and 97msTE with larger voxel size (2x2x2 cm<sup>3</sup>) (c). It is evident the baseline differences from short to longer TE and a decreasing noise increasing the voxel size.

#### Bibliography

1. Yan H et al. N Engl J Med 2009; 360:765-73. 2. Choi C et al. Nat Med 2012; 18(4):624-29. 3. Pope WB et al. J Neurooncol 2012; 107(1):197-205. 4. Andronesi OC et al. Sci Transl Med 2012; 4(116):116ra4. 5. Bertolino N et al. Phys Med. 2014 Sep;30(6):702-07. 6. Ledford H. Nature. 2014 Apr 10;508(7495):158-9 7. Provencher SW. Magn Reson Med. 1993; 30(6):6727-29. 8. Kalinina J et al. J Mol Med (Berl) 2012; 90(10):1161-71. 9. Andronesi OC et al. J Clin Invest. 2013 Sep 3;123(9):3659-63.

### Sin

#### XLVI CONGRESSO NAZIONALE 10-13 OTTOBRE 2015 – GENOVA

## WebPoster



